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SYNTHESIS OF POTENT β -D-GLUCOCEREBROSIDASE INHIBITORS: N-ALKYL- β -VALIENAMINES

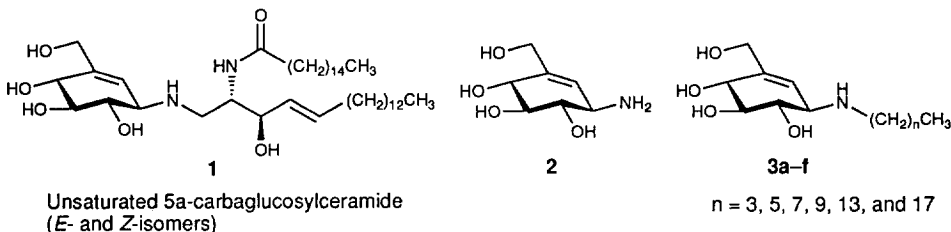
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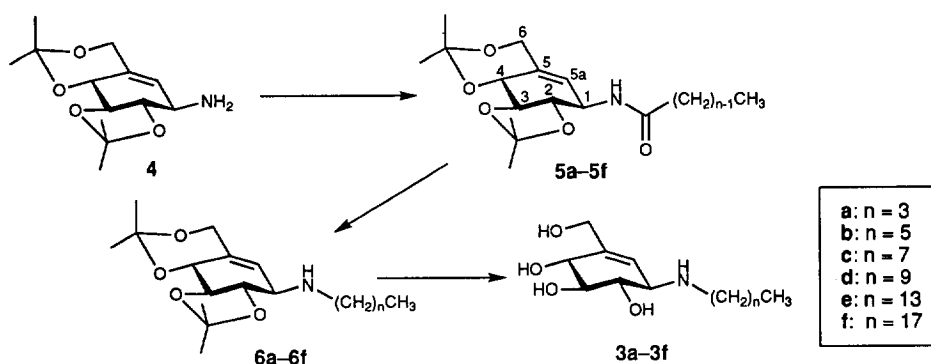
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Abstract: Six homologous derivatives (*N*-butyl **3a**, hexyl **3b**, octyl **3c**, decyl **3d**, tetradecyl **3e** and stearyl **3f**) of β -valienamine were synthesized. All have been shown to be potent and specific inhibitors of β -glucocerebrosidase, and to have no potency against glucosylceramide synthase (mouse liver microsomes). Among them, the *N*-octyl derivative possesses the strongest activity (IC_{50} 3×10^{-8} M), being almost 10-fold more potent compared to the unsaturated 5a-carba-glucosylceramide **1**. Compounds **3b** and **3c** are also moderate inhibitors of α -glucosidase (Baker's yeast). Copyright © 1996 Elsevier Science Ltd

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Recent findings^[1] of the potent and specific β -D-glucocerebrosidase inhibitors, 5a-carba- β -D-xylo-hex-5(5a)-enopyranosylceramides **1** (*E*- and *Z*-isomers) have stimulated us to develop a similar type of inhibitors by transforming β -valienamine **2** into some derivatives with more simple structures. Extensive studies^[2] on





specific glucosylceramide synthase inhibitor PDMP (D-*threo*-1-phenyl-2-decanoylamino-3-morpholino-1-propanol) and homologous series of compounds have so far been carried out in order to extend these potential from biological tools to therapeutic agents. Although 5a-carba-sugar analogues^[1] of glycosylceramides were initially designed as inhibitors of glycosyl transferase, they have been shown to possess strong inhibitory-activity against β -glucocerebrosidase, but no potency against glucosylceramide synthase at all. Therefore, it has also become of interest to elucidate the structural features of these ceramide-analogues, which would specifically differentiate the inhibitory action against the two kind of enzymes.

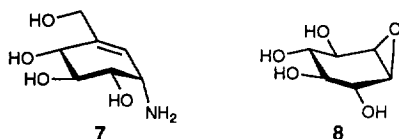
In this paper, in order to know inhibitory potency of 5a-carba-sugar analogues that mimic the potent glucosidase inhibitors^[3] *N*-alkyl glucopyranosylamines, six homologous *N*-alkyl β -valienamines **3a-3f** were synthesized by the conventional procedure: conversion of the protected β -valienamine **4** into the corresponding amides, reduction with lithium aluminum hydride, and subsequent deprotection. Thus, acylation of di-*O*-isopropylidene- β -valienamine^[4] (**4**) with *n*-octanoyl chloride (1.2 molar equiv.) in pyridine at room temperature produced the amide **5c**, quantitatively. Treatment of **5c** with an excess of lithium aluminum hydride (15 molar equiv.) in THF for 2 h at reflux temperature gave the protected amine **6c** in 85% yield. *O*-Deisopropylidenation was carried out with aqueous 80% acetic acid at 80°C for 4 h to give, after chromatography on silica gel with chloroform/methanol (3:1) as an eluent, the amine^[5] **3c** as acetate in quantitative yield. Other homologous series of compounds^[6] **3a**, **3b**, **3d**, **3e**, and **3f** were similarly prepared as in the preparation of **3c**.

Inhibitory activity^[7] of six compounds **3a-f** prepared in this work, along with those of *E*-1, conduritol B epoxide^[3] **8**, and α -**7** and β -valienamines **2**, were listed in Table 1. Inhibitory activity against β -glucocerebrosidase seems to be largely depend on the length of *N*-alkyl chain. The activity of the most

Table 1. Inhibitory Activity of Six *N*-Alkyl- β -valienamines and Its Related Compounds against Three Enzymes

Compound	Inhibitory activity (IC ₅₀ , M)		
	β -Glucocerebrosidase	Glucocerebroside synthase	α -Glucosidase
5a-Carba-glucosylceramide <i>E</i> -1	3.0×10^{-7}	NI	NT
β -Valienamine 2	NI	NI	1.0×10^{-4}
<i>N</i> -Butyl- β -valienamine 3a	1.1×10^{-5}	NI	*
<i>N</i> -Hexyl- β -valienamine 3b	3.0×10^{-7}	NI	5.0×10^{-5}
<i>N</i> -Octyl- β -valienamine 3c	3.0×10^{-8}	NI	1.7×10^{-5}
<i>N</i> -Decyl- β -valienamine 3d	7.0×10^{-8}	NT	NT
<i>N</i> -Tetradecyl- β -valienamine 3e	1.2×10^{-7}	NT	NT
<i>N</i> -Stearyl- β -valienamine 3f	3.0×10^{-7}	NI	*
α -Valienamine 7	NI	NI	1.0×10^{-4}
Conduritol B epoxide 8	1.1×10^{-6}	NI	*

*Activity less than IC₅₀ 1.0×10^{-4} M; NI: No inhibitory activity observed at 1.0×10^{-4} M; NT: Not tested



potent *N*-octyl compound **3c** has been shown to be stronger than those of the analogues *E*-1 and *Z*-1 containing the natural ceramide hydrophobic portions and that of conduritol B epoxide^[3] **8**, a well known irreversible inhibitor for this enzyme. These results demonstrated that the hydrophobic ceramide moiety can be replaced by a simple aliphatic chain without affecting the inhibitory activity, suggesting that this enzyme does not recognize so strictly the hydrophobic parts of the inhibitors. Since β -**2** and α -valienamines **7** have no activity against β -glucocerebrosidase, the *N*-alkyl chain with an appropriate length should be needed for exhibiting inhibitory-potency. Therefore, it is interesting to know the structure and inhibitory activity relationship of this kind of inhibitors by modifying the alkyl chains. Moreover, *N*-alkyl- β -valienamines are expected to act as specific competitive inhibitor, since valienamine moiety is seemed to mimic the transition state of the substrate for β -glucosidase reaction. On the other hand, compounds **3a–3f** completely lack inhibitory activity against glucocerebroside synthase. Then, our next targeted compounds should be the PDMP analogues, the morpholine parts of which being replaced by the β -valienamine residues.

References and Notes

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- [5] Physical properties of compounds: compound **5c**, [α]_D²⁴ -84° (*c* 0.8, CHCl₃), IR: 3280, 1650 cm⁻¹ (amide); ¹H NMR (270 MHz, CDCl₃): δ 5.56 (d, 1 H, *J* 7.7 Hz, NH), 5.40 (s, 1 H, H-5a), 4.71 (dd, 1 H, *J* 7.7, 9.5 Hz, H-1), 4.63 (ds, 1 H, *J* 8.12 Hz, H-4), 4.45 and 4.16 (ABq, *J* 14.3 Hz, H₂-6), 3.79 (dd, 1 H, *J* 8.1, 9.5 Hz, H-3), 3.57 (t, 1 H, *J* 9.5 Hz, H-2), 2.20 (ddd, 1 H, *J* 5.1, 7.7, 14.7 Hz, H-2'), 2.15 (ddd, 1 H, *J* 5.1, 7.7, 14.7 Hz, H-2'), 1.63 (m, 2 H, H₂-3'), 1.55, 1.46, and 1.42 (3 s, 3, 6, 3 H, 2 CMe₂), 1.28 (m, 8 H, H₂-4', 5', 6', 7'), 0.87 (t, 3 H, *J* 6.6 Hz, Me). **6c**, [α]_D²³ -46° (*c* 1.6, MeOH), IR: 3320 cm⁻¹ (NH); ¹H NMR (270 MHz, CDCl₃): δ 5.46 (s, 1 H, H-5a), 4.62 (d, 1 H, *J* 7.7 Hz, H-4), 4.50 and 4.15 (ABq, *J* 13.9 Hz, H₂-6), 3.73 (dd, 1 H, *J* 7.7, 9.5 Hz, H-3), 3.50 (d, 1 H, *J* 8.4 Hz, H-1), 3.49 (dd, 1 H, *J* 8.4, 9.5 Hz, H-2), 2.72 (ddd, 2 H, *J* 5.1, 7.7, 14.3 Hz, H₂-1'), 1.56, 1.46, 1.42 (3 s, 3, 6, 3 H, 2 CMe₂), 1.30 (m, 12 H, H₂-2', 3', 4', 5', 6', 7'), 0.87 (m, 3 H, Me). **3c**, [α]_D²² -39° (*c* 0.6, MeOH), IR: 2920, 2858, 1630 cm⁻¹ (NH₃⁺); ¹H NMR (270 MHz, CDCl₃/CD₃OD 2:1) δ 5.62 (s, 1 H, 5a-H), 4.22 and 4.17 (ABq, *J* 14.6 Hz, H₂-6), 4.16 (d, 1 H, *J* 7.7 Hz, H-4), 3.62 (t, 1 H, *J* 9.2 Hz, H-2), 3.58 (dd, 1 H, *J* 7.7, 9.2 Hz, H-3), 3.54 (d, 1 H, *J* 9.2 Hz, H-1), 2.99 (ddd, 1 H, *J* 7.5, 11.7 Hz, H-1'), 2.82 (ddd, *J* 7.5, 11.7 Hz, H-1'), 1.98 (s, 3 H, AcO⁻), 1.66 (m, 14 H, H₂-2', 3', 4', 5', 6', 7'), 0.89 (t, 3 H, *J* 6.6 Hz, Me).
- [6] All other compounds described herein gave the spectral data consistent with the assigned structures.
- [7] Inhibitory activities (IC₅₀) listed in Table 1 were measured by the following procedures.
 β -Glucocerebrosidase: The assay was performed with the fluorogenic substrate, NBD-glucosylceramide, with microsomal fraction of mouse liver in a total volume 0.2 ml containing glucocerebroside from Gaucher spleen, polyoxyethylene octylphenyl ether, sodium taurocholate and phosphate-citrate (pH 5.5) as reported^[2].
Glucosylceramide synthase: UDP-glucose:ceramide glucosyltransferase was assayed with liver microsomes with slight modification of the method of Inokuchi and Radin (Inokuchi, J.-i.; Radin, N. S. *J. Lipid. Res.* **1987**, *28*, 565). Liposomes were prepared from *N*-octanoylsphingosine, dioleoyl phosphatidylcholine and brain sulfatide. The mixture (liposome and microsome) was incubated for 1 h with UDP-[³H]glucose, β -NAD, DTT, EDTA, MgCl₂, and Tris-Cl (pH 7.4). The labeled GlcCer formed was isolated by partitioning between *t*-butyl methyl ether and 2-propanol aqueous Na₂SO₄ and counted without removing the precipitated protein.
 α -Glucosidase: The assay was performed with *p*-nitrophenyl α -D-glucopyranoside (0.66 mM) as substrates in 0.1 M phosphate buffer, pH 6.8, with α -glucosidase from Baker's yeasts reported (Uchida, C.; Kimura, H.; Ogawa, S. *Bioorg. Med. Chem. Lett.* **1994**, *22*, 2643).

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